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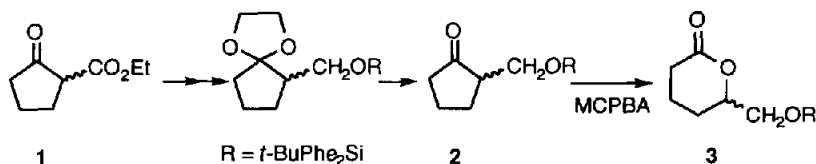
A Chemoenzymatic Preparation of Both Enantiomers of ω -Hydroxymethyl-Substituted Lactones

Didier Buisson and Robert Azerad

Laboratoire de Chimie et Biochimie Pharmacologiques et Toxicologiques associé au CNRS, Université René Descartes-Paris V, 45 rue des Saints-Pères, 75270-Paris Cedex 06, France

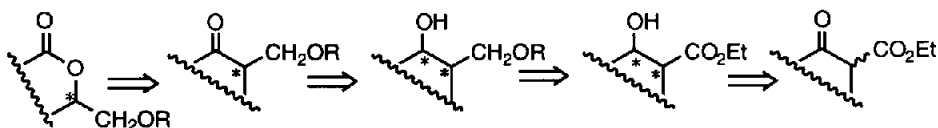
Abstract: (*R*)- and (*S*)- δ -hydroxymethyl valerolactone and ϵ -hydroxymethyl caprolactone were prepared as *tert*-butyldiphenylsilyl derivatives, in good yields and high enantiomeric purities, in a 5 step sequence, starting from the microbial stereospecific reduction of ethyl 2-oxocyclopentane or 2-oxocyclohexane carboxylates respectively.

Enantiomerically pure substituted lactones are important building blocks for a variety of natural biologically active products and pharmaceuticals. Of particular interest are protected 5-hydroxymethyl lactones¹⁻³, which have been prepared in racemic or homochiral forms, used for the synthesis of LTB₅¹, malyngolide⁴, and are potentially useful for the synthesis of various insect pheromones. Recently, an efficient and inexpensive synthesis of a protected (\pm)-6-hydroxymethyl-tetrahydro(2*H*)-pyran-2-one **3** has been reported⁵, via a regioselective Baeyer-Villiger reaction of the corresponding *tert*-butyldiphenylsilyl ether of (\pm)-2-hydroxymethylcyclopentanone **2**, itself obtained in three steps by acetal protection, reduction and deprotection, from ethyl 2-oxocyclopentane carboxylate **1**. Owing to the usually high stereoselectivity of the Baeyer-Villiger reaction^{6,7}, one may expect that the use of an enantiomerically pure suitably protected 2-hydroxymethylketone, in place of the racemic one, would result in an enantiomeric synthesis of the corresponding hydroxymethyl lactone.



Enantiomeric 2-hydroxymethyl cycloalkanones have been shown to be easily accessed by chemoenzymatic methods from (\pm)-cyclic β -oxoesters (Scheme 1). One method is based on a chemical reduction of the carbonyl group, followed by enzymatic resolution of one of the diastereomeric hydroxyesters^{8,9}. After reduction of the carboxyester group, which does not modify the stereochemistry at the α -carbon¹⁰, and protection of the resulting hydroxymethyl group, the secondary alcohol function may be oxidized back to the carbonyl group required for the Baeyer-Villiger oxidation¹¹. An even more attractive solution for obtaining in high yield a *single hydroxyester stereoisomer* is to replace the initial hydroxyester resolution by a microbial reduction^{12,13} of the corresponding (\pm)-cyclic β -oxoester, making use of the fast spontaneous epimerization of the substrate in the incubation medium, combined with the stereospecific reduction of a single enantiomer.^{10,14-17}. Such a diastereoselective and enantiospecific preparation of general-use cyclic synthons, having two

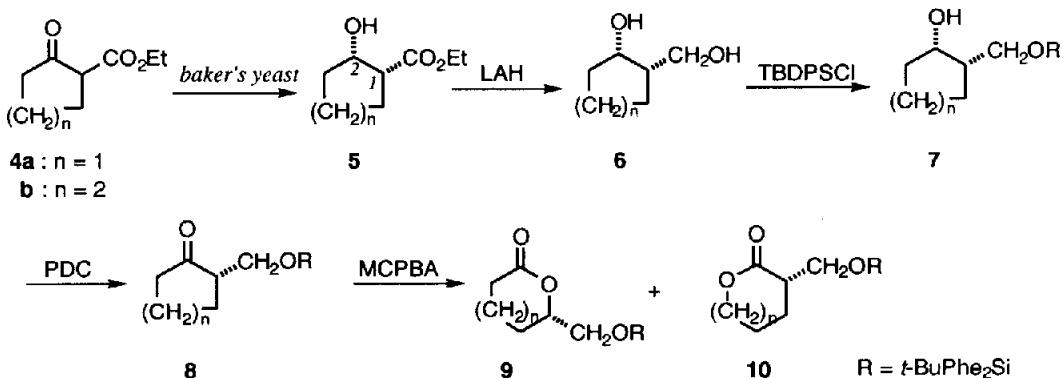
asymmetric centers, has been up to now moderately explored^{4,11,15,18-23}. The chemical counterpart of this method (so-called "dynamic kinetic resolution"), using catalytic hydrogenation in the presence of an asymmetric BINAP-Ru(II) complex, has been recently and concurrently developed²⁴⁻²⁷.



Scheme 1

We briefly describe herein the use of our microbial reduction method for obtaining, in good yields and high enantiomeric purities, both enantiomers of δ -hydroxymethyl valerolactone and ϵ -hydroxymethyl caprolactone.

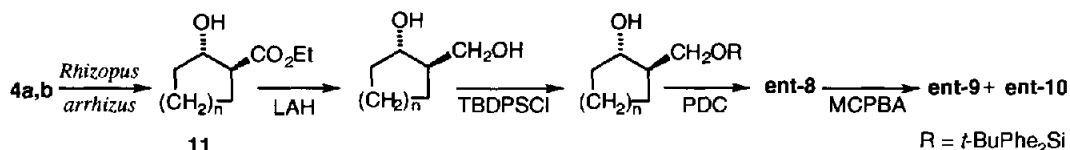
The reduction of ethyl 2-oxocyclopentanecarboxylate **4a** by baker's yeast¹⁴ (Scheme 2) afforded a major *cis* diastereomer (*cis/trans* ratio \geq 9:1) with a *1R,2S* configuration. After separation by flash chromatography, the *cis* hydroxyester **5a** (52% yield, 95% e.e.) was reduced with lithium aluminium hydride to give the (1*S*,2*S*)-diol **6a** (91% yield) which was protected with *tert*-butyldiphenylsilylchloride¹¹ to give the corresponding silyl derivative **7a** (87% yield). Oxidation with pyridinium dichromate in the presence of pyridinium trifluoroacetate²⁸ afforded the protected (*S*)-2-hydroxymethyl cyclopentanone **8a** (71% yield). In our hands, contrarily to the previously described reaction⁵, the oxidation of **8a** by *m*-chloroperbenzoic acid afforded a mixture of hydroxymethyl lactones **9a** + **10a** (16:1, 80% yield) which were separated by flash chromatography and characterized by ¹H- and ¹³C-NMR²⁹. The major, enantiomerically pure, oily (*S*)-5-*tert*-butyldiphenylsilyloxymethyl lactone **9a**, $[\alpha]_D^{20} +10$, $[\alpha]_{365}^{20} +34$ (c 0.7, CHCl₃), was thus obtained in 5 steps and a final 22% yield from the oxocarboxylate **4a**. A purified sample of the isomeric lactone **10a** showed $[\alpha]_D^{20} -18$, $[\alpha]_{365}^{20} -63$ (c 0.5, CHCl₃).



Scheme 2

The enantiomeric protected (*R*)-lactone **ent-9a**, $[\alpha]_D^{20} -11$, $[\alpha]_{365}^{20} -37$ (c 0.86, CHCl₃), was similarly obtained in a final 23 % yield, starting from the reduction of ethyl 2-oxocyclopentanecarboxylate **4a** by

*Rhizopus arrhizus*¹⁰ (Scheme 3), which afforded the *trans* enantiomerically pure (1*S*,2*S*)-hydroxyester **11a** (57% yield, after chromatographic separation from the *cis* isomer; initial *cis/trans* ratio, 2:8). The isomeric (*R*)-lactone **ent-10a** showed $[\alpha]_{\text{D}}^{20} +17$, $[\alpha]_{365}^{20} +59$ (c 0.63, CHCl₃).



It was attractive to apply such reactions to the synthesis of both enantiomers of the homologous 7-membered lactone, which might also constitute useful lactonic synthons. Baker's yeast^{10,14,15,19} (Scheme 2) allowed again the preparation of the nearly enantiomerically pure *cis* (1*R*,2*S*)-hydroxyester **5b** (80% yield, after chromatographic separation from a small amount of the *trans* isomer). Reduction to **6b** (92% yield), protection to **7b** (97% yield), PDC oxidation to **8b** (78% yield) and MCPBA oxidation, led to the exclusive formation of the protected (*S*)-hydroxymethyl lactone **9b**²⁹, m.p. 68-69° C, $[\alpha]_{\text{D}}^{20} -4$, $[\alpha]_{365}^{20} -17$ (c 1.3, CHCl₃), in a final 46% yield from the oxocarboxylate **4b**.

Similarly, using *Rhizopus arrhizus*¹⁰ (Scheme 3) for the reduction of **4b** to the enantiomerically pure *trans* (1*S*,2*S*)-hydroxyester **11b** (65% yield) and the same sequence of reactions, it was possible to obtain the enantiomeric (*R*)-lactone **ent-9b**, m.p. 69-70° C, $[\alpha]_{\text{D}}^{20} +4$, $[\alpha]_{365}^{20} +18$ (c 1.31, CHCl₃), in a final 32 % yield.

It is still possible to increase the convenience and the total yield of such conversions by effecting the separation of diastereomeric *cis* and *trans* derivatives, if necessary, at the protected diol stage, where a clear-cut separation is generally obtained, in place of the hydroxyester stage.

In conclusion, the easy availability of such versatile homochiral building blocks suggests that they will find a number of promising applications, as new members of the chiral pool, in the synthesis of a range of natural products. Work is in progress to extend this method to the stereospecific preparation of other lactonic synthons.

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29. All new compounds were fully characterized by spectroscopic data (¹H- and ¹³C-NMR, IR) and microanalysis and/or molecular ion mass measurements. Optical purities were determined by comparison with published optical rotations, when available, and/or by chiral GC or HPLC. Enantiomeric purities of lactones **9a-b** were determined by HPLC on a Chiralpak AD column (Daicel Chem. Ind., 250 x 4 mm, heptane-*i*PrOH 98:2, 1 mL.min⁻¹), and found to correlate, within the experimental error limits, with the enantiomeric excesses of hydroxyesters **5a-b** or **11a-b**.
9a, ¹H-NMR (250 MHz,CDCl₃) δ ppm, J Hz: 1.04 (9H, s, CH₃), 1.8 and 1.95 (4H, 2m, CH₂-4 and CH₂-5), 2.5 (2H, m, CH₂-3), 3.75 (2H, AB part of an ABX system, J_{AB}= 13, J_{AX}= 4.4, J_{BX}= 5.2, CH₂OSi), 4.37 (1H, m, H-6), 7.4 (6H, m, ArH), 7.65 (4H, m, ArH). ¹³C-NMR (62.9 MHz,CDCl₃) δ ppm: 171.13 (CO), 135.63, 135.56, 129.85, and 127.79 (ArCH), 133.06 and 132.88 (ArC), 80.15 (CH-6), 65.58 (CH₂O), 29.82, 24.38, and 18.24 (CH₂), 26.74 (CH₃), 19.20 (quat.CMe₃).
10a, ¹H-NMR (250 MHz,CDCl₃) δ ppm, J Hz: 1.04 (9H, s, CH₃), 1.85-2.3 (4H, 2m, CH₂-4 and -6), 2.65 (2H, m, CH-3), 3.88 (1H, dd, J_{3'a-3} = 3.6, J_{3'a-3'b} = 10, H-3'a), 4.02 (1H, dd, J_{3'b-3} = 6, J_{3'b-3'a} = 10, H-3'b), 4.32 (2H, m, CH₂-6), 7.39 (6H, m, ArH), 7.64 (4H, m, ArH). ¹³C-NMR (62.9 MHz,CDCl₃) δ ppm: 172.09 (CO), 135.70, 135.50, 129.85, 129.74 and 127.72 (ArCH), 133.42 and 133.00 (ArC), 71.54 and 64.26 (CH₂O), 42.75 (CH-6), 26.82 (CH₃), 22.84 and 22.26 (CH₂), 19.28 (quat.CMe₃).
9b, ¹H-NMR (250 MHz,CDCl₃) δ ppm, J Hz: 1.04 (9H, s, CH₃), 1.5, 1.95 and 2.18 (6H, 2m, CH₂-4, -5 and -6), 2.50 and 2.64 (2H, 2m, CH₂-3), 3.58 (1H, dd, J_{3'a-3} = 7, J_{3'a-3'b} = 10.3, H-3'a, CH₂OSi), 3.80 (1H, dd, J_{3'b-3} = 5.6, J_{3'b-3'a} = 10.3, H-3'b), 4.21 (1H, m, H-7), 7.4 (6H, m, ArH), 7.64 (4H, m, ArH). ¹³C-NMR (62.9 MHz,CDCl₃) δ ppm: 175.04 (CO), 135.61, 129.86, and 127.78 (ArCH), 133.19 (ArC), 80.20 (CH-6), 65.94 (CH₂O), 34.89, 31.06, 28.15 and 23.11 (CH₂), 26.86 (CH₃), 19.25 (quat.CMe₃).

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